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## Total synthesis of calonyctin A2, a macrolidic glycolipid with plant growth-promoting activity

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## Abstract

Calonyctin A2, a tetrasaccharidic glycolipid having a 22-membered macrolidic structure, has been synthesized by the assembly of three 6-deoxygenated thioglycoside intermediates. The short-step synthesis was achieved by preparation of the most complicated b–c disaccharide unit from phenyl 2,2':4,6:4',6'-tri-O-benzylidene-1-thio- $\beta$ -D-laminaribioside without any glycosidation reaction and by regioselective macrolactonization. © 2000 Elsevier Science Ltd. All rights reserved.

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Calonyctin extracted from the leaves of Yue-Guang-Hua (Calonyction aculeatum L. House) is a plant growth regulator, which promotes the tuber production of sweet potato and increases the crop yields of beans and wheat.<sup>1</sup> It is a mixture of homologous glycolipids consisting of a common deoxygenated tetrasaccharide residue and 11-hydroxy fatty acids,<sup>2</sup> which are named calonyctin A1 (1) and A2 (2), respectively (Fig. 1).<sup>3</sup> Furthermore, one of the sugar hydroxyl groups is acylated with (2R,3R)-3-hydroxy-2-methylbutyric acid. The most remarkable feature of their structure is that they have a 22-membered macloride ring. The absolute configuration of the aglycon moiety was determined as S by Schmidt's synthesis of **1** from a racemic fatty acid.<sup>4</sup> In this communication, we describe an expeditious synthesis of calonyctin A2 (2) using laminaribiose as the starting material. On the basis of our studies on the chemical modification of laminaribiose,<sup>5</sup> we envisioned that the readily accessible tri-O-benzylidene derivative **3** could be used as a synthon for the most complicated disaccharide unit (Qui-b and Qui-c) of 2. Compound **3** was converted into a crucial disaccharide donor, phenyl 3'-O-allyl-2,2'-di-O-benzoyl-4,4'-di-O-benzyl-6,6'-dideoxy-1-thio- $\beta$ -D-laminaribioside **8**, which had to be coupled with a monosaccharide intermediate 17 and a L-rhamnosyl donor 21. Furthermore, an optically active 11(S)-hydroxymyristic acid derivative 14 could be prepared from the known (S)- $\chi$ -tosyloxymethyl- $\chi$ -butyrolactone in an enantioselective manner.

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The synthetic route of the disaccharide donor **8** is illustrated in Scheme 1. Thus, **3** was allylated at the unprotected hydroxyl group to give the fully protected disaccharide<sup>6</sup> **4**. Upon treatment of **4** with PPTS in CHCl<sub>3</sub>–MeOH at room temperature, the most labile *O*-benzylidene group with an eightmembered ring underwent selective cleavage, to give the 2,2'-diol<sup>6</sup> **5** in 79% yield based on consumed **4**. The following reductive cleavage of the benzylidene group in **5** was one of the most difficult steps in the present synthesis, and this was overcome by applying Kusumoto's reagent system.<sup>7</sup> Treatment of **5** with BH<sub>3</sub>·Me<sub>2</sub>NH–BF<sub>3</sub>·OEt<sub>2</sub> in CH<sub>2</sub>Cl<sub>2</sub> gave the desired 2,6,2',6'-tetraol **6** in 77% yield without any affection on the other protecting groups.<sup>8</sup> The conventional reduction of **5**, or its 2,2'-di-*O*-acetyl derivative with BH<sub>3</sub>·Me<sub>3</sub>N–AlCl<sub>3</sub><sup>9</sup> or DAIBAL-H,<sup>10</sup> Super-Hydride<sup>®</sup>, failed due to poor regioselectivity and undesirable cleavage of the protecting group. Subsequently, **6** underwent selective esterification in a one-pot manner with TsCl and then with excess BzCl, giving the labile 2,2'-di-*O*-benzoyl-6,6'-di-*O*tosyl derivative **7**. Without purification, **7** was reduced with NaBH<sub>4</sub> in DMF at 70°C into the dideoxy derivative<sup>6</sup> **8** in an overall yield of 47% from **6**.



Scheme 1. Reagents and conditions: (a) AllylBr, NaH, DMF; (b) PPTS, MeOH/CHCl<sub>3</sub>; (c)  $BF_3 \cdot OEt_2$ ,  $Me_2NH \cdot BH_3$ ,  $CH_2Cl_2$ , under  $N_2$ ; (d) TsCl, pyridine, then BzCl; (e) NaBH<sub>4</sub>, DMF, 70°C

For the synthesis of the aglycon moiety of **2**, the optically active pentanetriol<sup>11</sup> **9** derived from Lglutamic acid was subjected to selective silvlation and subsequent basic treatment with *tert*-BuOK in THF to give the epoxide **10** (Scheme 2). Treatment of **10** in THF with 9-(benzyloxy)nonyl magnesium bromide in the presence of CuI and subsequent benzoylation gave **11**<sup>6</sup> in an overall yield of 72%. A three-step reaction involving de-*O*-silvlation with Bu<sub>4</sub>NF, bromination with CBr<sub>4</sub>–PPh<sub>3</sub>,<sup>12</sup> and reduction with Bu<sub>3</sub>SnH–AIBN was carried out to give **12**.<sup>6</sup> Treatment of **12** with methanolic sodium methoxide followed by Birch reduction gave the tetradecan-1,11-diol<sup>6</sup> **13**. Finally, a primary hydroxyl group of **13** was subjected to two-step oxidation<sup>13</sup> with TEMPO and subsequently with sodium chlorite to give 11(S)-hydroxymyristic acid, which was isolated and characterized as the methyl ester **14**.<sup>6</sup> In order to construct the target compound **2**, two other intermediates, phenyl 2-*O*-acetyl-3,4-di-*O*-benzyl-6-*O*-mesyl-1-thio- $\beta$ -D-glucopyranoside<sup>14</sup> **15** and phenyl 2,3,4-tri-*O*-benzyl-1-thio- $\alpha$ -L-rhamnopyranoside<sup>15</sup> **21**, were prepared.



Scheme 2. Reagents and conditions: (a) TBDPSCl, pyridine/CH<sub>2</sub>Cl<sub>2</sub>, then *tert*-BuOK, THF; (b) 9-(benzyloxy)nonyl magnesium bromide, CuI, THF, 0°C: BzCl, pyridine; (c) *n*-Bu<sub>4</sub>NF, THF: CBr<sub>4</sub>, Ph<sub>3</sub>P, DMF, then Bu<sub>3</sub>SnH, AIBN, toluene, 80°C; (d) NaOMe, MeOH, 70°C, then liq. NH<sub>3</sub>, Na, -80°C; (e) TEMPO, TBACl, NCS, CH<sub>2</sub>Cl<sub>2</sub>/H<sub>2</sub>O, pH 8.6: NaClO<sub>2</sub>, NaH<sub>2</sub>PO<sub>4</sub>, *tert*-BuOH, 2-methyl-2-butene/H<sub>2</sub>O, then CH<sub>2</sub>N<sub>2</sub>, Et<sub>2</sub>O; (f) thioglycoside<sup>14</sup> **15**, NIS–TfOH, CH<sub>2</sub>Cl<sub>2</sub>, MS 4 Å, -20°C; then NaOMe, MeOH; (g) NaBH<sub>4</sub>, DMF, 70°C; (h) **8**, NIS–TfOH, CH<sub>2</sub>Cl<sub>2</sub>, MS 4 Å, -20°C; (i) 2 M KOH, MeOH, 40°C; (j) Cl<sub>3</sub>C<sub>6</sub>H<sub>2</sub>COCl, Et<sub>3</sub>N, DMAP, toluene; (k) thioglycoside<sup>15</sup> **21**, NIS–TfOH, CH<sub>2</sub>Cl<sub>2</sub>, MS 4 Å, -20°C; (l) RhCl(PPh)<sub>3</sub>, diazabicyclo[2,2,2]octane, EtOH, reflux: 2 M HCl, 45°C, then (2*R*,3*R*)-3-benzyloxy-2-methylbutyric acid, DMAP, WSC, CH<sub>2</sub>Cl<sub>2</sub>; (m) Pd/C, H<sub>2</sub>, MeOH

Assembly of the four intermediates (8, 14, 15 and 21) were performed by iodonium activation of the thioglycosides. At first, 14 and the glucosyl donor 15 were treated with NIS–TfOH<sup>16</sup> in CH<sub>2</sub>Cl<sub>2</sub> at  $-20^{\circ}$ C, and the  $\beta$ -glucoside 16 was isolated after de-*O*-acetylation. Reduction of the mesyloxy group in 16 with NaBH<sub>4</sub> in DMF gave the glycosyl acceptor<sup>6</sup> 17, which was coupled with 8 in a similar way to afford the trisaccharide<sup>6</sup> 18 in 71% yield. After saponification with aqueous NaOH, the resulting carboxylic acid 19 was subjected to macrolactonization by a mixed anhydride procedure<sup>17</sup> under high-dilution conditions in toluene ( $1.71 \times 10^{-4}$  M). Although two possible hydroxyl groups are present at the 2b- and 2c-positions, the desired 22-membered macrolide<sup>6</sup> 20 was obtained as a single product in 60% yield together with recovered starting material 19. The structure of 20 was unambiguously corroborated by two-dimensional COSY and NOESY NMR spectroscopy. Similar glycosylation of 20 with the L-rhamnosyl donor 21 gave the tetrasaccharide<sup>6</sup> 22 in 15% yield, which was not improved by use of another thiophilic reagent, MeOTf. The low yield was probably due to steric hindrance of the hydroxyl

group at the 2a-position. To synthesize the target compound, the allyl group in **22** was isomerized with Wilkinson's catalyst and the propenyl group was removed by acid hydrolysis. The resulting alcohol was esterified with (2R,3R)-3-benzyloxy-2-methylbutyric acid<sup>4</sup> in the presence of water-soluble carbodiimide in CH<sub>2</sub>Cl<sub>2</sub>, giving the fully protected derivative **23** in 62% yield. Finally, removal of all benzyl groups in **23** by catalytic hydrogenolysis gave calonyctin A2 (**2**), of which the NMR spectra in pyridine- $d_5$  were consistent with those already reported.<sup>2</sup>

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- 6. All new compounds had satisfactory elemental analysis data or high-resolution mass spectra. Optical rotations were measured in CHCl<sub>3</sub> at 25°C unless otherwise noted. Selected data for 4: <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta_{\rm H}$  5.72, 5.59, 5.56 (each s, PhCH);  $[\alpha]_D$  -59.2 (c 2.0). Compound 5:  $\delta_H$  5.58, 5.53 (each s, PhCH), 4.68 (d, J=9.6, H-1), 4.65 (d, J=7.0, H-1');  $[\alpha]_D - 43.4$  (*c* 0.47 in CHCl<sub>3</sub>); HRMS (FAB+) *m*/*z* for C<sub>35</sub>H<sub>39</sub>O<sub>10</sub>S (M+H)<sup>+</sup>, calcd: 651.2263; found: 651.2236. Compound 6:  $\delta_{\rm H}$  5.72, 5.59, 5.56 (each s, Ph*CH*); [ $\alpha$ ]<sub>D</sub> -3.8 (*c* 5.74). Compound 8:  $\delta_{\rm H}$  4.75 (d, *J*=8.0, H-1'), 4.70 (d, J=10.1, H-1). Compound **9**:  $\delta_{\rm H}$  3.85 (m, CH), 3.65 (t, J=5.6, CH<sub>2</sub>). Compound **11**:  $\delta_{\rm H}$  3.69 (t, J=6.0, 14-CH<sub>2</sub>), 3.46 (t, J=6.7, 1-CH<sub>2</sub>), 2.00 (bs, OH); [α]<sub>D</sub> +2.1 (*c* 0.40 in CHCl<sub>3</sub>); HRMS (FAB+) *m*/*z* for C<sub>44</sub>H<sub>59</sub>O<sub>4</sub>Si (M+H)<sup>+</sup>, calcd: 679.4155; found: 679.4183. Compound **12**: δ<sub>H</sub> 0.93 (t, J=7.3, 14-CH<sub>3</sub>), 4.49 (s, PhCH<sub>2</sub>), 5.15 (m, 11-CH); [α]<sub>D</sub> +3.7 (c 0.29); HRMS (FAB+) m/z for  $C_{28}H_{41}O_3$   $(M+H)^+$ , calcd: 425.3056; found: 425.3060. Compound **13**:  $\delta_D$  3.64 (t, *J*=6.6, 1-CH<sub>2</sub>), 3.61 (m, 11-CH);  $[\alpha]_D$  +1.8 (*c* 0.80). Compound 14: m.p. 41.4–42°C;  $\delta_D$  3.67 (s, CH<sub>3</sub>O), 3.60 (m, 11-CH);  $[\alpha]_D$  0.7 (*c* 0.25). Compound **17**:  $\delta_{D}$  4.25 (d, J=7.3, H-1), 3.66 (s, COO*CH*<sub>3</sub>), 0.90 (t, J=7.1, CH<sub>3</sub>); [ $\alpha$ ]<sub>D</sub> -17.5 (c 5.44). Compound **18**:  $\delta_{D}$ 4.99 (d, J=7.5, H-1b), 4.73 (d, J=7.7, H-1c), 4.22 (d, J=7.5, H-1a); [α]<sub>D</sub> +10.1 (c 0.43). Compound **20**: δ<sub>D</sub> 4.99 (d, J=7.5, H-1a); H-1b), 4.73 (d, J=7.7, H-1c), 4.22 (d, J=7.5, H-1a);  $[\alpha]_D = 18.7$  (c 0.69); HRMS (FAB+) m/z for  $C_{63}H_{84}O_{14}Na$  (M+Na)<sup>+</sup>, calcd: 1087.5759; found: 1087.5740. Compound 22:  $\delta_{\rm H}$  ((CD<sub>3</sub>)<sub>2</sub>CO): 5.44 (d, J=0.9, H-1d), 4.32 (d, J=7.6, H-1a), 4.27 (bs, 1H, H-2d);  $[\alpha]_D = 6.3 (c \ 0.20)$ ; HRMS (FAB+) m/z for  $C_{90}H_{112}O_{18}Na (M+Na)^+$ , calcd: 1503.7746; found: 1503.7730. Compound 23: δ<sub>H</sub> ((CD<sub>3</sub>)<sub>2</sub>CO): 5.51 (t, J=9.4, H-3c), 5.41 (s, H-1d), 5.15 (d, J=7.9, H-1c), 5.08 (t, J=8.8, H-2c), 4.85 (d, J=7.8, H-1c), 5.08 (t, J=8.8, H J=8.8, H-1b), 4.38 (bs, H-2d), 4.31 (d, J=7.6, H-1a), 4.12 (bd, J=9.3, H-3d), 4.01 (t, J=9.4, H-3b), 3.61 (t, J=8.6, H-2a), 3.45 (t, J=8.4, H-4c), 3.37–3.27 (m, H-5a), 3.18 (t, J=8.8, H-4a), 3.21–3.11 (m, H-5a), 2.92 (t, J=9.1, H-4b), 2.33–2.22 (m,  $CH_2$ COO), 0.86 (t, J=7.0,  $CH_3$ );  $\delta_C$  (( $CD_3$ )<sub>2</sub>CO): 174.0, 172.8, 139.9, 139.4, 139.3, 139.2, 129.2, 128.7, 128.6, 128.5, 128.5, 128.6, 128.5, 128.5, 128.6, 128.5 128.3, 128.2, 128.1, 127.9, 127.8, 127.7, 127.6, 127.5, 103.0, 100.7, 99.6, 86.3, 84.4, 82.0, 81.9, 81.4, 78.8, 77.5, 75.8, 75.4, 74.9, 74.8, 74.7, 73.9, 72.8, 71.9, 71.6, 71.1, 45.3, 38.7, 35.6, 33.7, 31.0, 30.5, 30.3, 30.1, 29.8, 29.6, 29.3, 29.1, 28.8, 28.6, 25.9, 25.8, 25.4, 25.2, 22.8, 18.5, 18.2, 18.0, 17.9, 17.8, 15.9, 14.1, 13.9, 12.9, 12.1; [α]<sub>D</sub> - 8.9 (c 0.23); HRMS (FAB+) m/z for C<sub>99</sub>H<sub>123</sub>O<sub>20</sub>Na (M+H+Na)<sup>+</sup>, calcd: 1654.8505; found: 1654.8634. Compound **2**: [α]<sub>D</sub> –51.7 (*c* 0.12, EtOH); HRMS (FAB+) m/z for C<sub>43</sub>H<sub>74</sub>O<sub>20</sub>Na (M+Na)<sup>+</sup>, calcd: 933.4671; found: 933.4670.
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- 15. New compound **21** [<sup>1</sup>H NMR: 5.49 (bs, H-1);  $[\alpha]_D$  +0.65 (*c* 0.70)] was prepared by the Lewis acid-promoted thioglycosidation of the fully acetylated rhamnose, followed by de-*O*-acetylation and *O*-benzylation.
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